



## Bionew™ Plasmid DNA Purification Kit ( Medium Quantity )

**Size: 100 preps**

**#Cat: BN100-015a**

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### **I. Description**

Nucleic acid isolation is the most technically demanding and laborious procedure performed in molecular cloning laboratory. At first purification procedure usually used phenol, chloroform, etc. These hazardous organic chemicals, need multiple centrifugations and will result in significant loss of DNA, may affect the enzyme digestion of DNA or introduce amplification inhibitors.

Using spin column to purify DNA from samples is conventional method now, which required DNA was adsorbed to the membrane by centrifuge or vacuum, which bring not only DNA but also cell debris and other precipitate to the membrane. The membrane can be clogged because of the precipitate, then result in inefficient recovery of samples.

A recent developed DNA extraction method can directly use whole blood as the samples. It is the use of magnetic beads covered with nucleic acid-binding matrices. Its theory is nucleic acid (DNA and RNA) in the solution is specifically adsorbed to the surface of magnetic beads because the following action: (i) shielded intermolecular electrostatic forces, (ii) dehydration of the DNA/RNA and beads surfaces, and (iii) intermolecular hydrogen bond formation in the DNA/RNA-beads contact layer.

This method can farthest eliminate the inhibition in the plasmid DNA extraction course, because plasmid DNA in the lysate is specifically adsorbed to the surfaces of magnetic beads but the precipitate in the lysate isn't adsorbed. Then after 2 times of wash steps for the magnetic beads the other impurities are eliminated, finally the pure plasmid DNA is eluted and can be used in PCR or enzyme digestion directly.

Bionew™ plasmid DNA purification kit is the excellent kit which uses magnetic beads for plasmid DNA extraction. It can be widely used in plasmid DNA purification with sample from Research Institutions, Life Science and Industrial Markets. The procedures of plasmid DNA extraction system contains: lysis and DNA release, DNA binding to surface of magnetic beads, 2 times of wash and DNA elution.

For more information, see the [www.bionewtech.cn](http://www.bionewtech.cn) or contact Bionewtech Technical Service: [service@bionewtech.cn](mailto:service@bionewtech.cn) .

### **II. Components and Storage**

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Components	Size ( 100 preps )
Magnetic Beads Solution	1.1ml×2 tubes
Proteinase K(Lyophilized)	2 tubes
Proteinase K Solution Buffer	1.4ml×2 tubes
Lysis Buffer	22 ml×1 bottle
Wash 1 Buffer	22 ml×1 bottle
Wash 2 Buffer	22 ml×1 bottle
Elution Buffer	22 ml×1 bottle

Upon receipt, store all components at room temperature until Expiry Date.

Expiry Date: From Production Date, for up to 12 months.

### III. Applicability

The kit can be directly used for plasmid DNA purification from the cultured

#### . Materials Needed

Reagents: 100% Alcohol , 100% Isopropyl Alcohol

Expendable: Centrifuge Tube of 2ml, 1.5ml ; Tip of 200 $\mu$ l, 1000 $\mu$ l ; All sterilized by 1.01E+5Pa, 121 for 20 min. Recommend to use Axygen expendable.

Instruments: Pipettor(10 $\mu$ l,200 $\mu$ l,1000 $\mu$ l); Vortex Agitator, Constant Temperature Equipment (water bath or metal bath)

#### . Samples Pretreatment

Transfer the culture medium into 1.5ml centrifuge tubes, centrifuge at 4 with 10000g, 30s.

Discard the supernatant fully and entirely, and then use the precipitate for next purification.

#### . Protocol

##### (1) Preparation Protocol

- 1.1 For the tube of [Proteinase K (Lyophilized)], take 1.4 ml [Proteinase K Solution Buffer] into the tube and solved the [Proteinase K (Lyophilized)] by up and down the tube. It is the final [Proteinase K]. Then make aliquots of [Proteinase K] and store at 2~8 until expiry date.
- 1.2 For the bottle of 22ml [Wash 1 Buffer], add 11ml Alcohol, 11ml Isopropyl alcohol to the bottle and mix it by the bottle up and down, mark it to confirm the addition of alcohol and isopropyl alcohol. It is the final [Wash 1 Buffer]. Screw the cap closely and store at 2~8 for up to 6 months.
- 1.3 For the bottle of 22ml [Wash 2 Buffer], add 22ml Isopropyl alcohol to the bottle and mix it by the bottle up and down, mark it to confirm the addition of isopropyl alcohol. It is the final [Wash 2 Buffer]. Screw the cap closely and store at 2~8 for up to 6 months.
- 1.4 Before use vortex [magnetic beads solution] for 15s to mix equably. The dosage of beads is 20 $\mu$ l beads/sample, so for N samples take N×20 $\mu$ l beads into new centrifuge tube, then take N×200 $\mu$ l isopropyl alcohol into this tube, vortex the solution 15s to mix equably. It is the final [magnetic beads solution].
- 1.5 If the [Lysis buffer] forms a precipitate, warm the solution in 56 for 10mins to solve the precipitate completely.
- 1.6 For frozen samples take out to room temperature to unfreeze. Take out [Elution Buffer] in 56 to preheat it.

## (2) Purification Protocol

- 2.1. For N samples, use N centrifuge tubes (2ml) and separately marked it; add 25 $\mu$ l step1.1 [Proteinase K] firstly.
- 2.2. Add 100 $\mu$ l pretreated sample **from Part V.** using different 200 $\mu$ l tips
- 2.3. Add 100 $\mu$ l [Lysis Buffer] using 200 $\mu$ l tips, vortex for 5s, incubate the sample at 56 for 8 minutes.
- 2.4. Add 220 $\mu$ l step1.4 [Magnetic Beads Solution] using 200 $\mu$ l tips, vortex for 10s to mix it equably, incubate the sample at 56 for 3 minutes.
- 2.5. Place the N centrifuge tubes(2ml) containing sample mixture to the magnetic stand for 3 minutes, the magnetic beads are adsorbed to the wall of tube because of magnetic force, then discard all supernatant using different 1000 $\mu$ l tips.
- 2.6. Remove the tubes from the magnetic stand, add 400 $\mu$ l step1.2 [Wash 1 Buffer] using 1000 $\mu$ l tips, vortex 5s.
- 2.7. Place the tubes to the magnetic stand for 2 minutes, and discard all supernatant using 1000 $\mu$ l tips (permission of using the same 1000 $\mu$ l tip).
- 2.8. Remove the residual solution using 200 $\mu$ l tips.
- 2.9. Remove the tubes from the magnetic stand; add 400 $\mu$ l step1.3 [Wash 2 Buffer] into every tube with different 1000 $\mu$ l tips, mixed every sample by slowly drawing and pumping by 1000 $\mu$ l pipettor. Note: Don't let tips touch to residual solution on the upside of tube.
- 2.10. Use N new centrifuge tubes(2ml), marked as step 2.1, transfer every mixture of step 2.9 into every new marked 2ml centrifuge tube
- 2.11. Place the tubes to the magnetic stand for 2 minutes, and discard all supernatant using 1000 $\mu$ l tips (permission of using the same 1000 $\mu$ l tip).
- 2.12. Open the lids of tubes and dry at room temperature for 3minutes, and then remove again the residual solution at the bottom of tubes with 200 $\mu$ l tips
- 2.13. Add 56 preheated [Elution Buffer] 100 $\mu$ l (optional volume from 50 $\mu$ l -200 $\mu$ l), mixed every sample by slowly drawing and pumping by 200 $\mu$ l pipettor, incubate at 56 for 3 minutes.
- 2.14. Place the tubes to the magnetic stand for 2 minutes,
- 2.15. Transfer the supernatant to the new centrifuge tubes. It is the eluted genomic DNA, it can be used for next detection at once or stored at 4°C for 1 week or stored at -20 for 6 months.



2.16.The yield and purity can be determined by spectrophotometer. For double stand DNA:

$$\text{dsDNA Concentration} = (\text{OD}_{260} - \text{OD}_{320}) \times 50 \mu\text{g/ml}$$

$$\text{dsDNA Purity} = (\text{OD}_{260} - \text{OD}_{320}) / (\text{OD}_{280} - \text{OD}_{320}),$$

When using the frozen culture medium as the sample, its concentration and purity is decreased comparing with fresh sample, so the range of purity of pure dsDNA is between 1.7 and 2.0.

#### . Troubleshooting

For questions not addressed here, please contact bionewtech.

Web site: [www.bionewtech.cn](http://www.bionewtech.cn)

Email: [service@bionewtech.cn](mailto:service@bionewtech.cn)

Symptoms	Possible Causes	Comments
Poor yield	Too little sample used , or the initial amount of DNA in the sample is poor.	Increase initial amount of sample.
	The sample is frozen for a long time, and the DNA has been degraded	Use fresh samples or use more samples.
	Elution Buffer amount is not enough.	Be sure 100μl Elution is used to elute the DNA.
Beads in final elution	Beads is occasionally transferred by rapid pipetting	Vortex or mix solution, place in the magnetic stand and transfer eluate to new tube again.
Inconsistent yield	Inconsistent amounts of magnetic beads	Be sure to vortex the magnetic beads solution before use and vortex beads in the binding step.
	Inconsistent sample is used	Be sure to mix the sample equably before use.
Purity lower than 1.7		If the purity is in the range of 1.6~1.7, the sample can be used for next detection.
	The sample is frozen too long time.	If the purity is lower than 1.6, don't recommend use such sample to do next detection.

#### . Composition of Buffers and Solutions

Elution Buffer: 10mM Tris. (PH7.2)

#### .Related Products

Bionew™ Whole Blood DNA Purification Kit	#Cat: BN100-011a
Bionew™ Tissue DNA Purification Kit	#Cat: BN100-012a
Bionew™ DNA Micro Purification Kit	#Cat: BN100-014a
Bionew™ Plasmid DNA Purification Kit (Medium Quantity)	#Cat: BN100-015a
Bionew™ Bacteria DNA Purification Kit	#Cat: BN100-016a
Bionew™ Viral DNA Purification Kit	#Cat: BN100-031a
Bionew™ Viral RNA Purification Kit	#Cat: BN100-032a
Bionew™ Viral DNA/RNA Purification Kit	#Cat: BN100-033a
Bionew™ Cells Total RNA Purification Kit	#Cat: BN100-021a
Bionew™ Tissue Total RNA Purification Kit	#Cat: BN100-022a
Bionew™ Whole Blood Total RNA Purification Kit	#Cat: BN100-023a
Bionew™ Lockable Magnetic Stand	#Cat: BN200-011a